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# Liquid-liquid extraction of a recombinant protein, cytochrome $b_5$ , with aqueous two-phase systems of polyethylene glycol and potassium phosphate salts

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#### Abstract

The partitioning of cytochrome  $b_5$  in aqueous two-phase systems of polyethylene glycol (PEG) and potassium phosphate salts was investigated. Cytochrome  $b_5$  partitioning is enhanced with decreasing polymer molecular mass and with increasing tieline length and pH. The effect of cytochrome  $b_5$  mutation, by substitution of the glutamic acid at positions 56 and 92 of the polypeptide chain by a lysine, on protein partitioning was also studied. Partitioning of cytochrome  $b_5$  mutants shows the same dependence on tieline length and pH, following the order cytochrome  $b_5 >$  mutant 56 > mutant 92.

# 1. Introduction

The downstream processing of biological materials requires separation and purification techniques leading to a high degree of purification and high recoveries and to low operating costs. One of the bioseparation processes that fulfils these criteria is liquid-liquid extraction using aqueous two-phase systems.

This technique can be used in the early steps of the purification process (*e.g.*, separating proteins from cell debris), replacing difficult solidliquid separations, and also for further purification [1]; the scale-up is relatively simple because it utilizes equipment common in the chemical industry. Aqueous two-phase systems contain a high proportion of water in both phases, providing an excellent environment for cells, cell organelles or biologically active proteins [2]. In order to find suitable conditions for the extraction of proteins with aqueous two-phase systems, it is necessary to know the partitioning behaviour of the desired proteins. Protein partitioning depends on the physico-chemical parameters of the systems, namely the type and molecular mass of polymers, tieline length (a function of the concentration of the system components), type and concentration of component or added salts, pH, temperature and addition of biospecific affinity ligands [2].

This paper reports the partitioning of cytochrome  $b_5$  in polyethylene glycol (PEG)-potassium phosphate salt two-phase systems. The effects of polymer molecular mass, tieline length and pH on cytochrome  $b_5$  partitioning were investigated. The influence of cytochrome  $b_5$ mutation, by substitution of the glutamic acid at positions 56 or 92 of the polypeptide chain by a lysine, on protein partitioning was also studied, in order to evaluate the effect of genetic manipu-

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lation techniques on the downstream processing of proteins.

## 2. Experimental

# 2.1. Chemicals

PEG 400, 1000, 3350 and 8000 were supplied by Sigma and anhydrous potassium dihydrogenphosphate ( $KH_2PO_4$ ) and dipotassium hydrogenphosphate ( $K_2HPO_4$ ), both of analyticalreagent grade, by Merck.

#### 2.2. Cytochrome $b_5$

The cytochrome  $b_5$  used consists of the hydrophilic domain of cytochrome  $b_5$  from mouse [3] that was cloned in a pUC 13 vector and expressed in *Escherichia coli* (TB1) [4]. This protein has a molecular mass of 13 600 and an isoelectric point (pI) of 4.4 [5].

This protein was mutated by site-specific mutagenesis in the glutamic acid at positions 56 or 92 of the polypeptide chain that was substituted by a lysine [5]. The pI of the mutants is 4.7 [5].

Cytochromes  $b_5$  were produced by fermentation and purified using sequential chromatographic steps [5].

#### 2.3. Preparation of aqueous two-phase systems

Aqueous two-phase systems of total mass 6 g were prepared by weighing appropriate amounts of concentrated solutions of PEG,  $KH_2PO_4$  and  $K_2HPO_4$  solutions, which were added until the required pH value was obtained, and water into 10-ml graduated centrifuge tubes. The contents of the tubes was intensively mixed on a vortex agitator and then 100  $\mu$ l of purified cytochrome  $b_5$  solution (2.8 mg ml<sup>-1</sup>) were added. After vortex mixing, the phases were separated by centrifugation (10 min at 128 g).

The concentrations of cytochrome  $b_5$  and cytochrome  $b_5$  mutants in each phase were determined by measuring the absorbance at 411 nm (molar absorptivity  $\varepsilon = 130$  l mmol<sup>-1</sup> cm<sup>-1</sup> [4]) using a Hitachi UV-Vis spectrophotometer.

Table 1 PEG-salt systems used in cytochrome  $b_5$  partitioning studies

Polymer	PEG-potassium phosphates (%, w/w)		
	Tieline 1	Tieline 2	Tieline 3
PEG 400	16.7/14.8	17.7/15.7	19.7/17.7
PEG 1000	16.2/14.3	17.7/15.7	19.7/17.7
PEG 3350	14/11.8	17.7/15.7	19.7/17.7
PEG 8000	14/11.8	-	-

The pH in each phase was measured with a Metrohm pH meter.

The PEG-salt systems prepared are shown in Table 1.

#### 3. Results and discussion

The effect of polymer molecular mass, tieline length and pH on cytochrome  $b_5$  partitioning in PEG-potassium phosphate systems is shown in Fig. 1. The partition coefficient,  $K_p$ , was defined as the ratio between cytochrome  $b_5$  concentration in the upper and lower phases and the yield, Y, as the ratio between cytochrome  $b_5$  mass in the top phase and in the total system. Each  $K_p$ and Y value represents the average of at least two measurements.

Cytochrome  $b_5$  partition coefficients and yields increase with decrease in polymer molecular mass. For aqueous two-phase systems of PEG 400 cytochrome  $b_5$  is mainly in the PEG-rich phase under all the experimental conditions studied ( $K_p > 2.7$  and Y > 86%); for PEG 1000, depending on tieline length and pH, cytochrome  $b_5$  may partition preferentially to the salt-rich phase or to the PEG-rich phase; for PEG 3350 cytochrome  $b_5$  is mainly in the salt-rich phase except for the longest tieline at high pH ( $K_p =$ 1.8 and Y = 61%); for PEG 8000, with only one tieline tested, cytochrome  $b_5$  accumulates in the salt-rich phase at all pH values ( $K_p < 8 \cdot 10^{-3}$  and Y < 0.6%).

The effect of polymer molecular mass can be attributed to the incréasing number of hydrophilic end groups on shorter PEG chains, which reduces the overall hydrophobicity [6], and to



Fig. 1. Effect of polymer molecular mass, tieline length and pH on cytochrome  $b_5$  partition coefficient,  $K_p$ , and yield, Y, in PEG-potassium phosphate systems at 23°C. (a) No formation of two phases; (b) KH<sub>2</sub>PO<sub>4</sub> did not dissolve.

the excluded volume effects that increase with increasing polymer molecular mass [7].

The enhancement of cytochrome  $b_5$  partitioning with increase in tieline length (Fig. 1) may be attributed to changes in the relative composition of the phases [8]. An increase in tieline length promotes an increase in phosphate concentration in the lower phase whereas in the upper phase it remains relatively constant and equal to its solubility limit in PEG. This results in the salting-out of proteins from the phosphate-rich phase to the PEG-rich phase mediated by the excluded volume effects of PEG. In the limit proteins will precipitate.

Cytochrome  $b_5$  partitioning also increases with increase in pH (Fig. 1). This is probably due to

an increase of the  $[HPO_4^{2-}]/[H_2PO_4^{-}]$  ratio, which promotes the shift of the phase diagram to lower polymer and salt concentrations. It is well known that small multivalent anions such as  $HPO_4^{2-}$ , used in conjunction with PEG, are more effective in inducing phase formation than monovalent anions [9] owing to the conflicting interaction between ether oxygens of PEG and small ions of high charge density [10]. The approach of salts, with these multivalent anions, to the polymer surface is constrained and a region of salt-depleted structured water is created at the polymer surface which might permit protein polyanion-PEG interactions [10]. The increase in the protein charge with increase in pH seems to be less important because cytochrome  $b_5$  already has a high negative charge at the studied pH values.

Cytochrome  $b_5$  partitioning in systems of PEG 1000 is strongly affected by the tieline length and pH. By changing the pH and/or the tieline length of the two-phase systems it is possible to manipulate cytochrome  $b_5$  partitioning to the phosphate-rich phase or to the PEG-rich phase. This versatility of PEG 1000 systems for cytochrome  $b_5$  partitioning makes them very attractive for separation and purification of cytochrome  $b_5$  from an impure extract.

PEG 1000 was chosen for further partitioning studies with cytochrome  $b_5$  mutants. Fig. 2 shows the effect of tieline length and pH on the partition coefficients and yields of cytochrome  $b_5$ and cytochrome  $b_5$  mutants in PEG 1000-potassium phosphate systems. Partitioning of the modified proteins (mutants 56 and 92) also increases with increasing tieline length and pH. The partition coefficients and yields obtained follow the order cytochrome  $b_5 >$  mutant 56 > mutant 92.

The isoelectric point of cytochrome  $b_5$  mutants (pI 4.7) is greater than the isoelectric point of cytochrome  $b_5$  (pI 4.4), hence a decrease in modified protein partitioning would be expected due to the increase in their net positive charge. The differences in the partitioning behaviour of mutants 56 and 92 were attributed to the position of the mutated amino acids in the polypeptide chain. For mutant 92 the mutated amino acid is



Fig. 2. Effect of tieline length and pH on the partition coefficients,  $K_p$ , and yields, Y, of cytochrome  $b_5$  and cytochrome  $b_5$  mutants (56 and 92) in PEG 1000-potassium phosphate systems at 23°C. (b) KH<sub>2</sub>PO<sub>4</sub> did not dissolve.

situated at the C-terminus and is more accessible to interaction with both phases of the two-phasic system.

#### 4. Conclusions

Cytochrome  $b_5$  partitioning in PEG-potassium phosphate two-phase systems is enhanced with decreasing polymer molecular mass and increasing tieline length and pH. Partitioning of cytochrome  $b_5$  mutants using PEG 1000 systems show the same dependence on tieline length and pH, following the order cytochrome  $b_5 >$  mutant 56 > mutant 92. Owing to their versatility, PEG 1000-potassium phosphate systems seem to be suitable for application in more impure media.

The results of this work suggest that it is possible to change the partitioning behaviour of proteins by engineering the protein surface, for example by making it more positive or negative.

## 5. Acknowledgement

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